

Synthesis of the Macrocyclic Spermidine Alkaloid (\pm)-(2*R**,3*R**)-3-Hydroxycelacinnine

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Dedicated to *Edgar Heilbronner* on the occasion of his 80th birthday

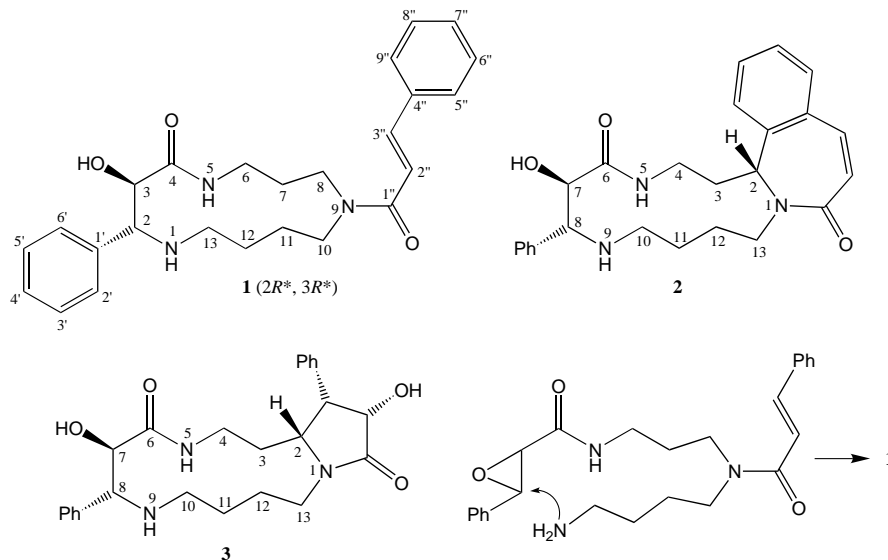
The macrocyclic lactam alkaloid (\pm)-(2*R**,3*R**)-3-hydroxycelacinnine (**1**) derived from spermidine was synthesized *via* stereoselective epoxide-ring opening with magnesium azide and cesium carbonate promoted macrocyclization of the ditosylated diamino precursor **12** with 1,4-dibromobutane in the two key steps (*Scheme 2*). ¹H- and ¹³C-NMR Signal assignments from COSY, HSQC, and HMBC 2D NMR data of the synthesized **1** were compared with the earlier-described data of the natural 3-hydroxycelacinnine. The similarity of their ¹³C-NMR spectra point to the correctness of the proposed constitutional formula for natural 3-hydroxycelacinnine; however, different ¹H-NMR chemical shifts and coupling constants ($J(2,3) = 9.0$ vs. 1.2 Hz, resp.) in the α -hydroxy- β -amino lactam moiety suggest that natural 3-hydroxycelacinnine is the 2,3-*cis*-epimer of one synthetic (\pm)-**1**.

Introduction. – Macrocyclic lactams derived from polyamines are of particular interest as synthetic targets for organic chemists due to their structural complexity and broad biological activity [1][2]. *Séguineau et al.* have isolated several novel hydroxylated spermidine alkaloids from the leaves of a New Caledonian Celastraceae, *Pleurostyliya opposita* (WALL.) MERRILL-METCALF [3][4]. Their proposed structures for 3-hydroxycelacinnine (**1**) (originally named 7-hydroxycelacinnine [3]), 7-hydroxypleurostyline (**2**), and 7-hydroxypleurocorine (**3**) are shown in *Scheme 1*¹⁾. The presence of an OH group at the α -position to the lactam carbonyl group represents a new feature in such alkaloids. A biosynthetic pathway of their formation involving an epoxy precursor has been suggested [3][4] (*Scheme 1*)²⁾. We are interested in the structure verification and biosynthesis of these alkaloids.

In this paper, we report the stereoselective synthesis of (\pm)-**1**. A large difference between coupling constants (9.0 vs. 1.2 Hz) as well as a significant difference between ¹H-NMR chemical shifts in the H–C(2)–C(3)–H moiety of the synthesized **1** and the natural 3-hydroxycelacinnine [3][4] suggest that the proposed relative *trans*-configuration (2*R**,3*R**) for the natural alkaloid should be changed to the corresponding *cis*-configuration (2*R**,3*S**). Also, the same conclusion holds for **2** and **3**, as well, since all three natural alkaloids had almost identical ¹H- and ¹³C-NMR data for this moiety [3][4].

- ¹⁾ To have the same atom numbering for **1–3**, *Séguineau et al.* proposed the name 7-hydroxycelacinnine for **1** [3]. However, in this paper, we use the name 3-hydroxycelacinnine according to the IUPAC rules for atom numbering, as shown in *Scheme 1*. For systematic names, see *Exper. Part*.
- ²⁾ In the light of the results discussed below, the alleged precursor for natural 3-hydroxycelacinnine should be the (*Z*)-epoxide.

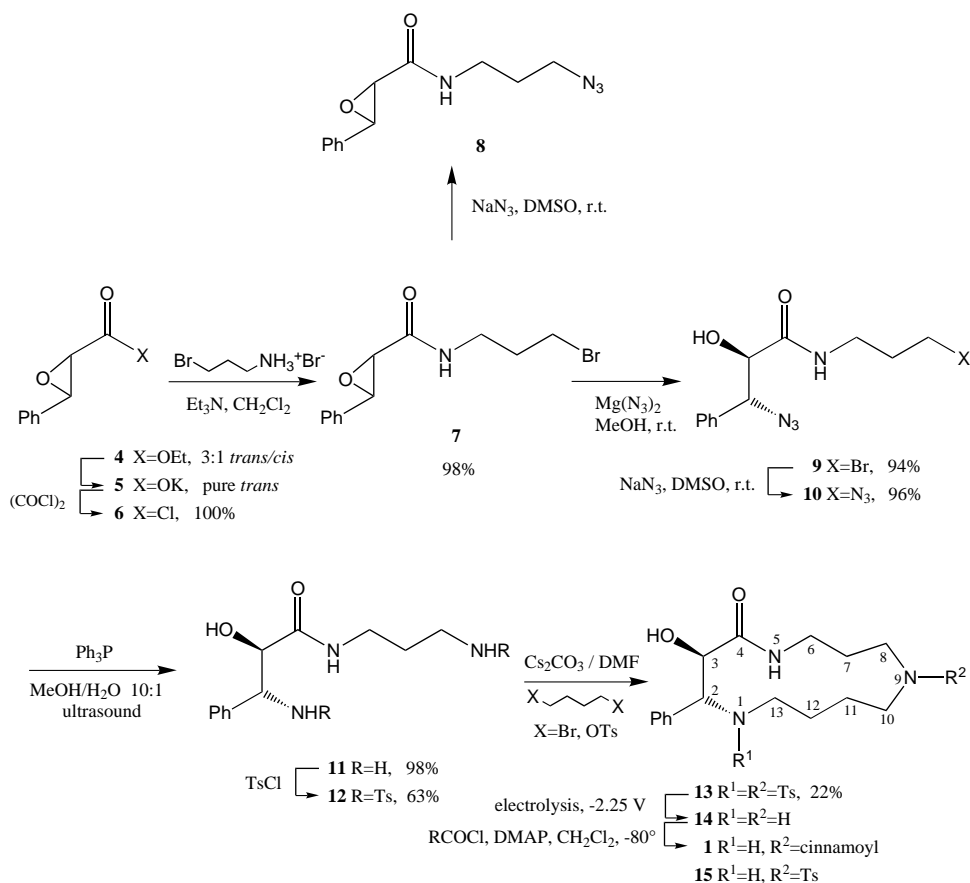
Scheme 1. Proposed Structures of Novel Hydroxylated Alkaloids 3-Hydroxycyclacinnine (**1**)¹⁾, 7-Hydroxypleurostyline (**2**), and 7-Hydroxypleurocorine (**3**) Isolated from *Pleurostylia opposita* and the Proposed Biosynthetic Formation of **1** [3][4]



Synthesis. – For the synthesis of **1**, the commercially available *ca.* 3 : 1 mixture of the *cis*- and *trans*-ethyl phenylglycidate (=ethyl 3-phenyloxiranecarboxylate; **4**) was saponified to its potassium salt [5], which gave pure potassium *trans*-oxiranecarboxylate **5** after crystallization from aqueous EtOH, as described for the sodium salt [6] (Scheme 2). Potassium carboxylate **5** was converted to its acid chloride **6** [7] according to the procedure described for the sodium salt. Coupling of **6** with 3-bromopropanamine hydrobromide promoted by Et₃N in CH₂Cl₂ provided the amide **7** in 98% yield from **5**.

Direct coupling of the bromo epoxy amide **7** with deprotonated TsNH₂ led to a complex product mixture containing only traces of the ditosylate **12**. Thus, longer routes from **7** to **12** had to be investigated. We found that the epoxide or bromide functionalities in **7** can be selectively cleaved by the azide ion under specific conditions keeping the other functionality intact. Following the procedure of *Behrens* and *Sharpless* for the stereo- and regioselective preparation of β -azido- α -hydroxy-amides [8][9], we obtained the monoazido derivative **9** in 94% yield by stirring a MeOH solution of **7** with an *in situ* (MgSO₄ + NaN₃) preparation of Mg(N₃)₂. In this particular case, the Ph-substituent at the β -position greatly facilitated the epoxide opening, and the reaction proceeded to a full completion after 2 h at room temperature. An attempted direct formation of the diazido derivative **10** from **7** required higher temperatures, which led to the formation of by-products. Also, the formation of the C(α) ring-opened isomer was below detectable level, which has to be compared to the 10 : 1 regioselectivity observed in the case of epoxyamides with aliphatic substituents after reflux in methanolic Mg(N₃)₂ for several hours, the conditions applied in the original work of *Behrens* and *Sharpless* [9]. The facile conversion of azidobromo-

Scheme 2



amide **9** into diazido derivative **10** was achieved in 96% yield by treating the former with an excess of NaN_3 in DMSO at room temperature for 2 h according to a described general procedure [10]. Under these conditions, bromoepoxyamide **7** can be selectively converted into azido epoxy amide **8**. Keeping the reaction mixture at room temperature for a longer time (several days) led to only partial epoxide cleavage of **8**, with the formation of diazido derivative **10** as well as several unidentified by-products, according to the ^1H - and ^{13}C -NMR analysis of the reaction mixture in (D_6) DMSO .

The general procedure for the facile azide reduction with Ph_3P in aqueous THF at room temperature [11] required a longer reaction time (> 24 h) in the case of diazido derivative **10** and furnished a low yield of diaminoamide **11**. A temperature increase to 50° facilitated the reduction. However, a small amount of by-product was always detected due, perhaps, to the slow reduction of the more hindered azido group at the secondary C(3) atom and the presence of the vicinal $\text{OH}-\text{C}(2)$, which may lead to several by-products [11]. Therefore, the reaction was studied by ^1H -NMR in various

aqueous deuterated solvents. The reduction of diazido derivative **10** with Ph_3P in $\text{MeOH}/\text{H}_2\text{O}$ 10 : 1 was best achieved by sonication of the suspension for 1 h to give **11** in 98% yield. For comparison, the reduction of **10** in aqueous DMSO under the same conditions proceeded much slower and with the formation of by-products.

Diaminohydroxyamide **11** was then selectively converted into ditosylate **12** with TsCl (2 equiv.), promoted by Na_2CO_3 in aqueous dioxane, in 63% yield after recrystallization. Tosylation of **11** in CH_2Cl_2 in the presence of Et_3N also led to **12** contaminated with a small amount of tritosylated product.

Several bimolecular [12–16] and intramolecular [17–20] macrocyclization reactions have been described based on the alkylation of *N*-monosubstituted sulfonamides promoted by Cs_2CO_3 in DMF [12–17] or K_2CO_3 in MeCN [18][19], or under *Mitsunobu* conditions [20]. According to previous studies, macrocyclization [12][13] as well as alkylation of tosylamides in general [13] proceeded better when $\text{Cs}_2\text{CO}_3/\text{DMF}$ was used, and high dilution was not usually required to achieve the best yields. With this method, we already have obtained several 13-membered spermidine-derived lactam alkaloids in up to 90% yield [21][22]. Thus, we prepared the ditosylated lactam **13** by Cs_2CO_3 -promoted coupling between **12** and a 1,4-disubstituted butane derivative. To evaluate effectiveness of this method for the preparation of similar macrocyclic lactams, we investigated the macrocyclization reaction between **12** and 1,4-dibromobutane under various conditions (see *Table 1* and *Scheme 3*).

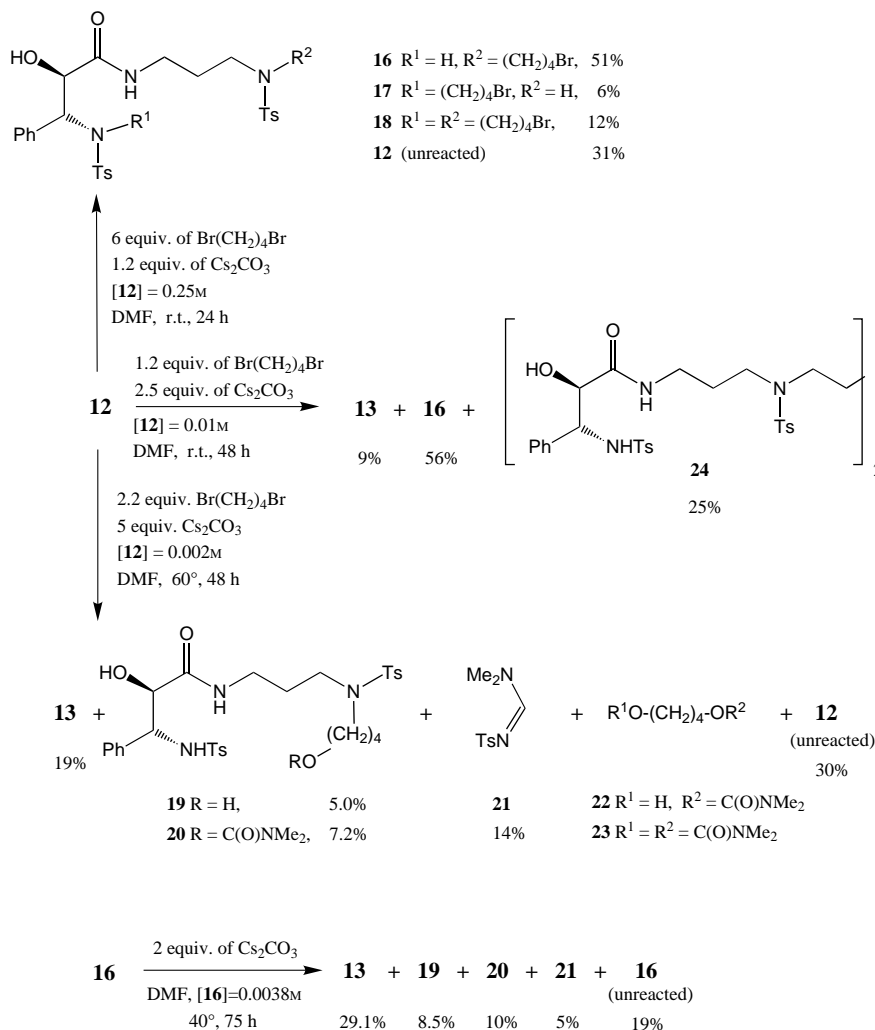
First, the reaction was run at room temperature with a slight excess of 1,4-dibromobutane (1.2 equiv.) and Cs_2CO_3 (2.5 equiv.) at moderate dilution ($[\mathbf{12}] = 0.01\text{M}$) (*Table 1, Entry 1*). Macrocyclic **13** was isolated in only 9% yield after 48 h together with the less hindered monosubstituted **16** (56%) as the major product and **24** (25%). The latter resulted from alkylation of the less hindered N-atoms of two molecules of **12** with one molecule of 1,4-dibromobutane. Presumably, **13** was formed from the more hindered monoadduct **17**, which was not detected in the reaction mixture due to its facile cyclization into **13**.

Reaction of **12** with an excess of 1,4-dibromobutane (6 equiv.) and an insufficient amount of Cs_2CO_3 (1.2 equiv.) in the minimum amount of DMF at room temperature (*Entry 2*) showed a *ca.* 10:1 selectivity between the two tosylated N-atoms of **12**

Table 1. Isolated Yields of **13** after Cs_2CO_3 -Promoted Macrocyclization of **12** under Various Conditions

Entry	Concentration of 12 [M] ^{a)}	$\text{X}(\text{CH}_2)_4\text{X}^b$ [equiv.]	Cs_2CO_3 [equiv.]	<i>T</i> [°C], time	13 [%]
1	0.01	1.2	2.5	r.t., 48 h	9
2 ^{c)}	0.2	6	1.2	r.t., 22 h	0
3 ^{c)}	$[\mathbf{16}] = 0.0038$	–	2.4	40°, 75 h	29
4 ^{c)}	0.002	1.1 + 1.1	2.1 + 1.1	60°, 48 h	19
5 ^{c)}	0.015	1.1	2.5	50°, 24 h	22
6	0.01	1.2 (X = OTs)	2.5	r.t., 72 h; then 60°, 64 h	15
7	0.075 (DMSO)	1.2	2.5	50°, 24 h; then r.t., 96 h	6.5
8	0.004 (DMSO)	2	2.5	40°, 72 h	< 10 ^{d)}
9	0.004 (MeCN)	2	2.5	40°, 72 h; then 60°, 72 h	22

^{a)} Solvent = DMF, unless noted otherwise. ^{b)} X = Br, unless noted otherwise. ^{c)} See text and *Exper. Part* for the detailed procedure and product distribution. ^{d)} Only traces of **13** were observed by TLC after workup, and the yield was estimated without separation.

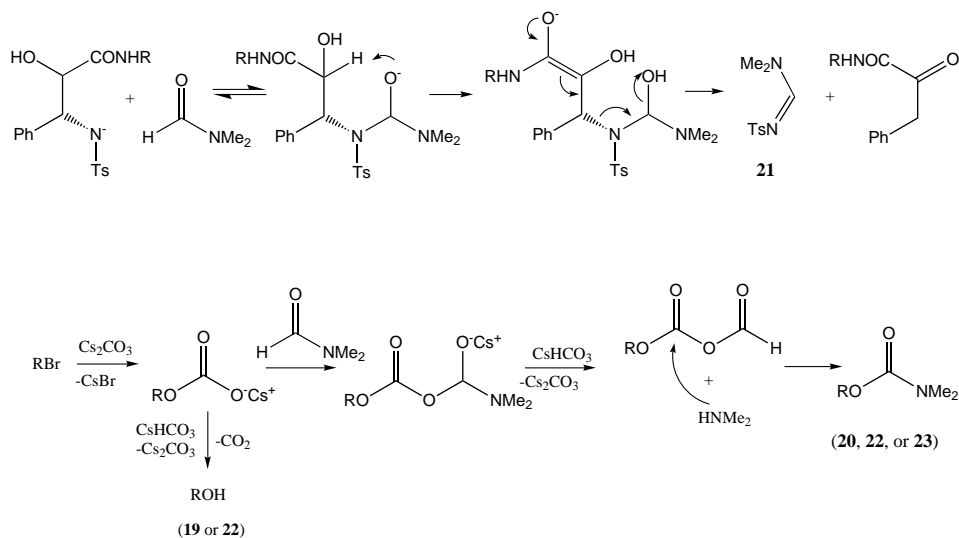
Scheme 3. Product Distribution after Cs_2CO_3 -Promoted Alkylation of **12** under Various Conditions

towards alkylation. Thus, the less hindered monoadduct **16** was again the major product (51%) accompanied by only 6% of the more hindered **17**, dialkylated **18** (12%), and unreacted **12** (31%). The isolated monoadduct **16** was then allowed to react with Cs_2CO_3 at moderate dilution ($[\text{16}] = 0.0038\text{M}$, *Entry 3*). No intramolecular macrocyclization was observed at room temperature. However, a very slow macrocyclization took place at 40° . After 75 h, lactam **13** was obtained in 29% yield along with unreacted **16** (19%), hydroxy compound **19** (8.5%), carbamate **20** (10%), formimidamide **21** [23] (5%), and several other unidentified products, presumably formed by β -elimination of tosylamide and elimination of bromide according to NMR analysis of the crude fractions obtained after column-chromatography separation.

At higher dilution ($[12] = 0.002\text{M}$, *Entry 4*), intermolecular monoalkylation of **12** by 1,4-dibromobutane occurred very slowly. Thus, a higher temperature (60°) and an excess of 1,4-dibromobutane (2.2 equiv.) were used to accelerate the reaction. However, several by-products were isolated (*Scheme 3*), similar to those obtained by macrocyclization of **16** after 48 h. The desired lactam **13** was isolated in only 19% yield, along with unreacted **12** (30%), hydroxy compound **19** (5%), carbamate **20** (7%), formimidamide **21** (14%), and carbamates **22** and **23**.

Thus, intramolecular macrocyclization of intermediate **16** involving the more hindered N-atoms proceeded slowly, with a rate comparable to the rates of side reactions, including DMF-assisted β -elimination of tosylamide with the formation of formimidamide **21** (*Scheme 4*), and solvolysis of the monobromo compound **16** to the hydroxy compound **19** and carbamate **20**. Carbamate **20** was presumably formed from **16** according to the mechanism depicted in *Scheme 4*. Although an excess (2.2 equiv.) of 1,4-dibromobutane was used, no dialkylated **18** was detected under high-dilution conditions, and no **24** was observed. The best yield of **13** (22%, *Entry 5*) in the bimolecular macrocyclization was achieved under moderate dilution conditions (0.015M) at 50° in DMF with 1.1 equiv. of 1,4-dibromobutane.

Scheme 4. Proposed Mechanisms of Side Reactions during Macrocyclization



The same by-products were isolated when ditosylated 1,4-butanediol was used in the macrocyclization instead of 1,4-dibromobutane, but no yield improvement was achieved (*Entry 6*). To avoid DMF-assisted side reactions, the macrocyclization was also investigated in DMSO and MeCN. In DMSO (*Entries 7* and *8*), the yields of **13** were even smaller, with a significant amount of hydroxy compound **19** being formed. Reaction in MeCN (*Entry 9*) with an excess of 1,4-dibromobutane (2 equiv.) at 40° proceeded slower than in DMF, with the formation of **16** as the major product, a small amount of the desired **13**, and a significant amount of the unreacted **12**. Temperature

increase to 60° led to only 22% yield of **13** after 72 h, due to the side reactions including, presumably, β -elimination of NHTs, elimination of bromide, and lactam cleavage.

Thus, we were unable to increase the yield of the macrocyclization step above 22%, and alternative routes to **1** were investigated [24]. According to the above-mentioned experimental data, the moderate yields in this step are best explained by the low reactivity of the more hindered tosylated N-atom towards alkylation, which allows slow side reactions to compete with the macrocyclization. With the less hindered ditosylated diamines studied by us, macrocyclization proceeded with somewhat better yields. In particular, with a substrate similar to **12** (with the β -phenyl group, but without the OH group), the optimized yield of the macrocyclization step was 45% [21]. With primary-alkyl moieties in the β -position, the macrocyclization proceeded with the highest yields [22]: the best yield (90%) was observed in the case of the unprotected hydroxymethyl substituent.

Finally, **13** was electrochemically detosylated at -2.25 V to give hydroxylactame **14** according to the procedure of *Guggisberg et al.* [25], followed by selective monoacylation with cinnamoyl chloride at the less hindered N-atom of the macrocycle [26] to give **1**.

NMR Data Analysis. – Complete ^1H - and ^{13}C -NMR signal assignments for the macrocyclic **1** and **13** were unambiguously obtained from the 2D NMR data (COSY, HMBC, and HSQC) and summarized in *Tables 2* and *3*. Due to the restricted rotation in the cinnamamide and very small differences in the chemical shifts of two rotamers, several ^1H - and ^{13}C -NMR signals of **1** gave broad lines at room temperature in both CDCl_3 and $(\text{D}_6)\text{DMSO}$. Thus, 2D NMR characterization of **1** was performed at the highest possible temperature before decomposition (70°, $(\text{D}_6)\text{DMSO}$) and also at low temperature (-40° , $\text{CDCl}_3/\text{CD}_3\text{OD}$ 30:1). However, even at 70°, the ^{13}C -NMR resonances of the four methylene C-atoms in the vicinity of the *N*-cinnamoyl group still appeared as broad doubled *s*. Also at low temperature, full resolution of all signals was not achieved, due to the very small chemical-shift difference between some resonances of two rotamers.

Unexpectedly, both H- and C-atoms of the β -amino- α -hydroxylactam fragment in ditosylated **13** gave broad signals at room temperature due, perhaps, to the restricted rotation around the crowded endocyclic TsN(1)–C(2)Ph bond. Thus, 2D NMR characterization of **13** was performed at 60° in $(\text{D}_6)\text{DMSO}$. Synthesized **1**, **13**, and **14** showed large coupling constants ($J(2,3) = 9.0\text{--}9.5$ Hz) for the two vicinal protons at C(2) and C(3), which were almost independent of solvent ($(\text{D}_6)\text{DMSO}$, CDCl_3) and temperature. Moreover, ^1H -NMR chemical shifts and proton coupling of the CH(2)–CH(3) moiety of the synthesized **1** and **14** were identical with those of **15** (*Table 3*) synthesized by a different route [24], but completely different from those of the natural 3-hydroxycelacinnine [3][4]. Based on these observations and the similarity of the rest of their ^1H -NMR spectra as well as on the general similarity of their ^{13}C -NMR spectra, we conclude that the natural 3-hydroxycelacinnine should have the reverse relative configuration at the two stereocenters as **1**. The natural alkaloids with the proposed [3][4] structures **1–3** gave also almost identical NMR data for the C(2)–C(3) fragment [3][4]; thus, the same correction of the relative configuration at this fragment should be applied to all three natural alkaloids. We are currently working

Table 2. Comparison of ^1H - and ^{13}C -NMR Data of Natural 3-Hydroxycyclacinnine [3][4] (2R*,3S*)-1^a, and Synthesized (2R*,3R*)-3-Hydroxycyclacinnine (1)

	(2R*,3S*)-1 [3] ^a		1 (this work) ^b		1 (this work) ^c	
	^1H	^{13}C	^1H	^{13}C	^1H	^{13}C
CH(2)	4.27 (<i>d</i> , $J(2,3) = 1.2$)	64.01, 64.38	3.62 (<i>d</i> , $J(2,3) = 9.1$)	65.79	3.64 (br. <i>d</i> , $J(2,3) = 8.9$)	66.08
CH(3)	4.16 (<i>d</i> , $J(2,3) = 1.2$)	76.07, 76.16	3.90 (<i>d</i> , $J(2,3) = 9.0$)	75.62	4.04 (br. <i>d</i> , $J(2,3) = 8.9$), 4.07 (br. <i>d</i> , $J(2,3) = 10$)	74.77, 74.41
C(4)		172.50, 172.34		173.13	–	174.11, 174.19
NH(5)	7.00 (<i>t</i>)		7.87 (br. <i>t</i>)		8.06, 8.27 (2 br. <i>s</i>)	–
CH ₂ (6)	3.80 (<i>m</i>), 3.00 (<i>m</i>)	36.72, 36.16	3.24 (<i>m</i>), 3.16 (<i>m</i>)	35.45	3.35 (<i>m</i> , 1 H), 3.39 (<i>m</i> , 1 H)	36.39, 35.95
CH ₂ (7)	2.25 (<i>m</i>), 1.70 (<i>m</i>)	27.92, 30.39	1.75–1.93 (<i>m</i>)	29.6 (br.), 27.8 (br.)	1.9–2.6 (br. <i>m</i> , 2 H)	27.17, 29.72
CH ₂ (8)	3.75 (<i>m</i>), 3.43 (<i>m</i>)	43.03, 44.59 ^d	3.63 (<i>m</i>), 3.28 (<i>m</i>)	44.1 (br.), 46.0 (br.)	3.68 + 3.58 ^e , 3.72 + 3.33 ^e	44.03, 47.08
CH ₂ (10)	3.57 (<i>m</i>), 3.38 (<i>m</i>)	44.01, 46.80 ^d	3.45–3.65 (<i>m</i>)	42.7 (br.), 43.2 (br.)	3.90 + 3.03 ^e , 3.54 + 3.59 ^e	44.8
43.38						
CH ₂ (11)	1.92 (<i>m</i>), 1.62 (<i>m</i>)	24.88 ^e , 25.12	1.76 (<i>m</i>), 1.56 (<i>m</i>)	25.8 (br.), 23.8 (br.)	1.66 + 1.78 ^e , 1.70 + 1.90 ^e	22.74, 25.27
CH ₂ (12)	1.65 (<i>m</i>), 1.45 (<i>m</i>)	24.22 ^e , 26.06	1.43 (<i>m</i>), 1.32 (<i>m</i>)	23.80	1.38–1.58 (2 br. <i>m</i> , 2 H)	23.75
CH ₂ (13)	2.86 (<i>m</i>), 2.31 (<i>m</i>)	46.55, 46.46	2.51 (<i>m</i>), 2.28 (<i>m</i>)	44.87	2.60 + 2.42 ^e , 2.64 + 2.40 ^e	44.62, 44.91
HO–C(3)			4.87 (br. <i>s</i>)		–	–
<i>Phenyl:</i>						
C(1')		142.35, 142.44		141.87	–	140.15, 139.92
CH(2',6')	7.35 (<i>m</i>)	127.76 ^f	7.34–7.37 (<i>m</i>)	127.45 ^e	7.38–7.49 ^d	128.48, 128.45 ^d
CH(3',5')	7.35 (<i>m</i>)	126.99 ^f	7.34–7.37 (<i>m</i>)	127.62 ^e	7.33–7.37 ^d	127.36 ^d
CH(4')	7.35 (<i>m</i>)	127.56 ^f	7.34–7.37 (<i>m</i>)	128.87	7.31–7.38	127.60, 127.53
<i>Cinnamoyl:</i>						
C=O	165.16, 165.82		164.56	–	166.12, 166.28	
CH=CHCO	6.30 (<i>d</i> , $J(2,3) = 15.4$)	117.53	7.05 (br. <i>d</i> , $J(2,3) = 15.6$)	118.80	6.83 (<i>d</i> , $J = 15.4$), 6.87 (<i>d</i> , $J = 15.4$)	116.30
CH=CHCO	7.66 (<i>d</i> , $J(2,3) = 15.4$)	170.79	7.48 (<i>d</i> , $J(2,3) = 15.4$)	140.34	7.69 (<i>d</i> , $J = 15.4$), 7.71 (<i>d</i> , $J = 15.4$)	142.80, 143.04
C(1'')		135.41	135.41	135.08	–	134.46
HC(2'',6'')	7.52 (<i>m</i>)	128.78	7.66 (br. <i>d</i> , $J = 7$)	127.45	7.57 (br. <i>d</i>), 7.59 (br. <i>d</i>)	127.75, 127.72
HC(3'',5'')	7.35 (<i>m</i>)	128.78	7.30–7.42 (<i>m</i>)	128.32	7.40–7.49	128.70, 128.68
HC(4'')	7.35 (<i>m</i>)	129.49	7.22 (<i>m</i>)	126.23	7.38–7.46	129.82, 129.76

^a) Configuration tentative. Spectra in CDCl_3 at r.t. with SiMe_4 as internal reference; ^1H and ^{13}C at 270 and 67.5 MHz, respectively. ^b) 18 mg in 0.6 ml of (D_6)DMSO, + 70°, $\delta(\text{DMSO})$ 2.50 (^1H , 300 MHz), and 39.51 (^{13}C , 75 MHz). ^c) 16 mg in 0.6 ml of $\text{CDCl}_3/\text{CD}_3\text{OD}$ 30 : 1, – 40°, $\delta(\text{SiMe}_4) = 0.00$ (^1H , 500 MHz), $\delta(\text{CDCl}_3)$ 77.00 (^{13}C , 125 MHz). ^d)^e)^f) Interchangeable values each. ^e) (2 br. *m*, 0.5 + 0.5 H).

Table 3. ^1H - and ^{13}C -NMR Data of Macroyclic **13**–**15**

	13 ^{a)}		14 ^{b)}		15 ^{c)}	
	^1H	^{13}C	^1H	^{13}C	^1H	^{13}C
CH(2)	5.19 (<i>d</i> , $J(2,3) = 9.5$)	63.62	3.56 (<i>d</i> , $J(2,3) = 9.0$)	66.62	3.47 (<i>d</i> , $J(2,3) = 10.1$)	66.06
CH(3)	4.72 (<i>d</i> , $J(2,3) = 9.6$)	70.97	3.98 (<i>d</i> , $J(2,3) = 9.0$)	73.78	3.99 (<i>d</i> , $J(2,3) = 10.1$)	72.61
C(4)		171.13		173.31		173.61
NH(5)	7.82 (br. <i>t</i> , $J = 5$)		7.13–7.28		8.51 (br. <i>t</i> , $J = 5$)	
CH ₂ (6)	3.38 (<i>m</i>), 3.08 (<i>m</i>)	36.29	3.43 (<i>m</i>), 3.28 (<i>m</i>)	39.84	3.53 (<i>m</i>), 3.25 (<i>m</i>)	37.36
CH ₂ (7)	1.84 (<i>m</i>), 1.74 (<i>m</i>)	27.79	1.68 (<i>m</i> , 2 H)	28.04 ^{d)}	1.90–1.98 (<i>m</i> , 2 H)	29.08
CH ₂ (8)	3.10 (<i>m</i>), 2.91 (<i>m</i>)	45.25	2.68–2.80 ^{d)}	49.69 ^{e)}	3.28 (<i>m</i>), 3.16 (<i>m</i>)	46.59
CH ₂ (10)	2.90 (<i>m</i>), 2.80 (<i>m</i>)	48.70	(<i>m</i> , 2 H)			
CH ₂ (11)	1.27 (<i>m</i> , 2 H)	24.43	2.43 ^{d)} (<i>m</i>), 2.26 ^{d)} (<i>m</i>)	49.00 ^{e)}	3.39 (<i>ddd</i>), 2.88 (<i>ddd</i>)	48.55
CH ₂ (12)	1.45 (<i>m</i> , 2 H)	25.51	1.53–1.68 ^{e)}	27.99 ^{d)}	1.83 (<i>m</i>), 1.69 (<i>m</i>)	24.20
CH ₂ (13)	3.05 (<i>m</i>), 2.96 (<i>m</i>)	46.11	(<i>m</i> , 3 H)			
HO–C(3)	–		1.53–1.68 ^{e)} , 1.36 ^{e)} (<i>m</i> , 1 H)	27.71 ^{d)}	1.48 (<i>m</i>), 1.41 (<i>m</i>)	25.29
			2.58 (<i>m</i> , 2 H) ^{d)}	45.97 ^{e)}	2.60 (<i>ddd</i>), 2.35 (<i>ddd</i>) 3.70 (br. <i>s</i>)	43.96
<i>Phenyl</i> :						
C(1'')		137.15		138.51		140.46
CH(2'',6'')	7.36 (<i>m</i>)	128.89	7.13–7.28	128.39 ^{f)}	7.23 (<i>d</i> , $J(2,3) = 7.26$, 2 H)	127.56
CH(3'',5'')	7.26 (<i>m</i>)	127.39	7.13–7.28	127.89 ^{f)}	7.37 (<i>t</i> , $J = 7.55$, 2 H)	128.91
CH(4'')	7.27 (<i>m</i>)	126.81	7.13–7.28	128.89	7.29 (<i>t</i> , $J = 7.4$, 1 H)	128.00
<i>1-Tosyl</i> ^{g)} :						
C(1')	–	137.55				
CH(2',6')	7.52 (<i>d</i> , $J(2,3) = 8.2$)	127.29				
CH(3',5')	7.18 (<i>d</i> , $J(2,3) = 8.2$)	128.59				
C(4')		142.40				
Me	2.31 (<i>s</i>)	20.55				
<i>9-Tosyl</i> ^{g)} :						
C(1')	–	134.89			–	136.32
CH(2',6')	7.63 (<i>d</i> , $J(2,3) = 8.2$)	126.73			7.68 (<i>d</i> , $J(2,3) = 8.2$)	127.40
CH(3',5')	7.41 (<i>d</i> , $J(2,3) = 8.2$)	129.40			7.32 (<i>d</i> , $J(2,3) = 8.3$)	129.95
C(4')	–	142.73			–	143.60
Me	2.40 (<i>s</i>)	20.55			2.43 (<i>s</i>)	21.69

^{a)} (D₆)DMSO, 60°, $\delta(\text{DMSO})$ 2.50 ppm (^1H , 300 MHz), and 39.51 (^{13}C , 75 MHz). ^{b)} CDCl₃, 23°, ^1H at 300 MHz, ^{13}C at 75 MHz, SiMe₄ as internal reference. No 2D NMR analysis was performed for this compound. ^{c)} CDCl₃, 27°, ^1H at 600 MHz, ^{13}C at 150 MHz, SiMe₄ as internal reference. ^{d)}^{e)}^{f)} Interchangeable values each. ^{g)} Aromatic protons of the tosyl group at N(1) gave 2 *d* (br.) in ^1H -NMR at r.t., which allowed unambiguous signal assignments for both Ts groups.

on the synthesis of 3-hydroxycelacinnine with the (2*R*,3*S*) absolute configuration which is most probably correct for the natural 3-hydroxycelacinnine. This assumption is based on the observation that all the macrocyclic spermine and spermidine alkaloids with known absolute configuration have the same relative configuration of the three substituents at C(2) (N-atom, phenyl or alkyl group, and substituted or unsubstituted acetamide moiety) [27].

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Experimental Part

General. All solvents were of anal.-grade quality and were used without further purification. DMF was stored under flame-dried molecular sieves 4 Å. DMSO and MeCN were dried and stored over CaH₂. Anh. MeOH under molecular sieves was purchased from *Fluka*. Ultrasound: *Branson B-220-125-W* ultrasonic cleaner. Flash chromatography (FC): silica gel *Merck 60* (40–63 μm, 230–400 mesh). TLC: *Merck* precoated silica-gel *60 F₂₅₄* plates; detection by UV at 254 nm, by 0.05% *Fluram* (*Fluka*) in acetone at 366 nm for primary amines, *Schlittler* reagent (H₂PtCl₆/HCl/KI) for amines and polyamines [28], Ce/Mo reagent (Ce(SO₄)₂·4 H₂O (10 g), aq. H₃[P(Mo₃O₁₀)₄] (25 g), H₂SO₄ (60 ml), H₂O (940 ml)) for epoxides and alcohols. M.p.: *Mettler FP-5/FP-52*. FT-IR: *Perkin-Elmer Spectrum One*, KBr pellets or neat/NaCl; in cm⁻¹. ¹H- and ¹³C-NMR: *Bruker ARX-300, DRX-500, or AMX-600*, ¹H at 300, 500, or 600 MHz and ¹³C at 75, 125, or 150 MHz, resp.; ¹³C {¹H}, DEPT-135, and DEPT-90 were recorded for all compounds; chemical shifts δ in ppm rel. to internal SiMe₄ (CDCl₃), unless noted otherwise, *J* values in Hz, nonobvious signal assignments by comparison with the spectra of the described similar compounds or by 2D NMR data; COSY, HMBC, and HSQC for macrocyclic products; all new compounds were at least 95% pure as judged by careful integration and peak analysis of ¹H-NMR spectra. EI-MS (70 eV) and CI-MS (NH₃ as reactant gas): *Finnigan-MAT 90*. ESI-MS (NaI/MeOH/CH₂Cl₂): *Finnigan TSQ-700* mass spectrometer; in *m/z* (% of base peak). Elemental analyses were performed by the Microanalytical Laboratory, Institute of Organic Chemistry, University of Zurich.

trans-3-Phenyloxiranecarbonyl Chloride (**6**) [7]. Oxalyl chloride (4.2 ml, 48.9 mmol) was added dropwise to a cold (ice/water bath) stirred suspension of potassium *trans*-phenylglycidate (**5**; 6.20 g, 30.69 mmol) in THF (90 ml) within 2 min. The cold bath was removed and stirring continued at 23° for 40 min until gas evolution ceased. The mixture was evaporated at r.t. to give a pale yellow oil, which solidified after 1 h under h.v.: 8.01 g of white solid as an equimolar mixture **6**/KCl, which was used in the next step without purification. M.p. 29–32°. FT-IR (KBr): 3050_w, 3036_w, 1778_s (C=O), 1600_w, 1497_w, 1458_m, 1408_m, 1289_w, 1237_w, 1196_w, 1117_m, 1085_m, 1058_w, 1012_m, 897_m, 852_w, 807_w, 751_s, 697_s, 663_w, 611_m, 583_w. ¹H-NMR (30 mg in 0.6 ml of CDCl₃, 300 MHz): 7.37–7.42 (*m*, 3 H); 7.28–7.33 (*m*, 2 H); 4.21 (*d*, *J* = 1.6, H–C(3)); 3.80 (*d*, *J* = 1.6, H–C(2)). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.91): 171.01 (COCl); 133.3 (C_{ipso}); 129.57 (C_p); 128.79 (C_m); 125.82 (C_o); 61.90 (C(2)); 59.13 (C(3)). ESI-MS: 201 (45, [RCO₂Me + Na]⁺), 233 (100, [PhCH(OMe) – CH(OH) – CO₂Me + Na]⁺); EI-MS: 147 (2, [M – Cl]⁺), 91 (100), 77 (40).

trans-N-(3-Bromopropyl)-3-phenyl-oxiranecarboxamide (**7**). Et₃N (12.8 ml, 92.1 mmol) in CH₂Cl₂ (60 ml) was added dropwise to a cold (ice/water bath) stirred suspension of freshly prepared **6** (7.966 g, 30.18 mmol) and 3-bromopropanamine hydrobromide (8.734 g, 39.9 mmol) in CH₂Cl₂ (60 ml) within 10 min. The mixture was stirred at 10° for 2 h and quenched with 100 ml of 5% aq. citric acid. The org. phase was washed with 5% citric acid (50 ml), H₂O (50 ml), and sat. aq. NaHCO₃ soln. (2 × 30 ml). Each aq. phase during these extractions was shaken with a small amount of CH₂Cl₂ and the obtained org. layer combined with the main org. soln. before the following extraction. The final org. phase was dried (Na₂CO₃) and evaporated to give a pale brown oil, which solidified under h.v. (9 h): pure **7** (8.55 g, 98.1% from **5**). Pale yellow solid. M.p. 83.5–85.5°. *R*_f (5% MeOH/CHCl₃) 0.8. FT-IR (KBr): 3271_s (NH), 3080_m, 2993_w, 2965_w, 2908_w, 2879_w, 2856_w, 1965_w, 1951_w, 1895_w, 1880_w, 1806_w, 1650_s (C=O), 1560_s, 1497_w, 1480_w, 1457_w, 1439_w, 1426_w, 1353_w, 1311_w, 1251_m, 1230_w, 1110_w, 1092_w, 1071_w, 1044_w, 1027_w, 997_w, 959_w, 884_m, 848_w, 823_w, 788_w, 769_w, 741_m, 720_w, 695_m, 673_m. ¹H-NMR (300 MHz, 30 mg in 0.7 ml of CDCl₃): 7.32–7.40 (*m*, 3 H); 7.23–7.29 (*m*, 2 H); 6.45 (br. *s*, NHCO); 3.88 (*d*, *J* = 2.0, H–C(3)); 3.52 (*d*, *J* = 2.0, H–C(2)); 3.37–3.51 (*m*, CH₂Br, CH₂N); 2.12 (*quint.*, *J* = 6.6, CH₂). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.93): 167.64 (CONH); 134.67 (C_{ipso}); 128.98 (C_p); 128.56 (C_m); 125.68 (C_o); 59.03 (C(2)); 58.83 (C(3)); 37.48 (CH₂NH); 31.98 (CH₂); 30.31 (CH₂Br). ESI-MS: 306, 308 (99, 100, [M + Na]⁺), 589, 591, 593 (4, 8, 4, [2M + Na]⁺). EI-MS: 283, 285 (2, 2, *M*⁺), 204 (3, [M – Br]⁺), 120 (30), 91 (100), 77 (38). Anal. calc. for C₁₂H₁₄BrNO₂ (284.15): H 4.97, C 50.72, N 4.93; found: H 4.98, C 50.68, N 4.76.

Reaction of 7 with NaN₃ in (D₆)DMSO: *trans-N-(3-Azidopropyl)-3-phenyl-oxiranecarboxamide* (**8**). The soln. prepared from **7** (52 mg, 0.183 mmol), (D₆)DMSO (0.7 ml), D₂O (0.05 ml), and NaN₃ (35.7 mg, 0.549 mmol) was kept at r.t. in the NMR tube and monitored by ¹H- and ¹³C-NMR. After 2 h, no starting **7** could be detected; the main product was **8** besides traces of diazidoamide **10** (*ca.* 3%; see below). After 3 days, **8/10** 2:1 and *ca.* 10% of by-products were observed after 6 days, **8/10** 2:3 and *ca.* 15% of by-products.

Data of 8: ¹H-NMR (300 MHz, δ((D₅)DMSO) = 2.55): 7.3–7.42 (*m*, 5 arom. H); 4.06 (*d*, *J* = 1.9, H–C(3)); 3.63 (*d*, *J* = 1.95, H–C(2)); 3.38 (*t*, *J* = 6.8, CH₂N₃); 3.21 (*t*, *J* = 6.8, CH₂NH); 1.72 (*quint.*, *J* = 7.8, CH₂). ¹³C-NMR (75 MHz, δ((D₆)DMSO) = 38.98): 166.78 (CONH); 135.38 (C_{ipso}); 128.61 (C_p); 128.43 (C_m); 125.84 (C_o); 57.71 (C(2)); 56.65 (C(3)); 48.25 (CH₂N₃); 35.75 (CH₂NH); 27.92 (CH₂).

(±)-(2R*,3R*)-3-Azido-N-(3-bromopropyl)-2-hydroxy-3-phenylpropanamide (**9**). Commercial anh. MgSO₄ (5.07 g, 42.25 mmol) was flame-dried and allowed to cool under N₂. Anh. MeOH (40 ml) and NaN₃ (2.75 g, 42.25 mmol) were introduced and sonicated for 5 min. A soln. of **7** (4.00 g, 14.08 mmol) in anh. MeOH (40 ml) was added at 0° and stirred under N₂. The reaction proceeded very slowly at 0°, but, at 23°, only traces of starting **7** were detectable after 60 min by TLC (SiO₂, 2.5% MeOH/CHCl₃). The mixture was stirred for a total of 2 h at 23° and quenched with H₂O (500 ml). The solid was filtered, washed with H₂O (4 × 50 ml), and briefly dried *in vacuo* to give ca. 4 g of wet product. The supernatant was extracted with CHCl₃ (4 × 100 ml), the extract combined with the solid fraction, dried (Na₂CO₃), and evaporated, and then the residue dried under h.v.: 4.313 g of **9** (93.6%), which contained less than 5% of impurities according to ¹H-NMR. The product was recrystallized from CHCl₃/hexane (10 ml of CHCl₃ for 1 g of **9**, followed by 20 ml of hexane): 3.8 g of pure **9** in two crops. White solid. M.p. 87–89°. FT-IR (KBr): 3300s, 3250s (NH, OH), 3033m, 3062m, 2966m, 2950m, 2921m, 2892m, 2851m, 2738w, 2494w, 2360w, 2220w, 2112s (N₃), 1957w, 1886w, 1807w, 1744w, 1634s (C=O), 1540s, 1494m, 1453m, 1426m, 1382w, 1372w, 1355m, 1305m, 1257s, 1196m, 1106s, 1086m, 1058w, 1028w, 991w, 964w, 949w, 935w, 913w, 871w, 838w, 813w, 788w, 741m, 725m, 696s, 658m, 639m. ¹H-NMR (300 MHz, 43 mg in 0.6 ml of CDCl₃): 7.37 (s, 5 arom. H); 6.48 (br. t, NHCO); 5.08 (d, *J* = 3.8, H–C(3)); 4.47 (br. d, *J* = 3.8, H–C(2)); 3.76 (br. s, OH); 3.37 (sext. (ddd), *J* = 6.7, 1 H, CH₂NH); 3.16 (q (ddt), 1 H, CH₂NH); 3.09 (dt, ³*J* = 6.4, ²*J* = 10.3, 1 H, CH₂Br); 2.94 (dt, ³*J* = 6.7, ²*J* = 10.3, 1 H, CH₂Br); 1.79 (quint., *J* = 6.5, CH₂). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.93): 169.95 (CONH); 133.99 (C_{ipso}); 128.86 (C_p); 128.45 (C_m); 128.28 (C_o); 73.95 (C(2)); 67.13 (C(3)); 37.23 (CH₂NH); 31.67 (CH₂); 30.15 (CH₂Br). CI-MS: 327, 329 (10, 10, [M + H]⁺), 247 (100, [M – Br]⁺), 204 (25, [M – Br – HN₃]⁺). Anal. calc. for C₁₂H₁₃BrN₃O₂ (327.18): H 4.62, C 44.05, N 17.12; found: H 4.92, C 44.11, N 17.51.

(±)-(2R*,3R*)-3-Azido-N-(3-azidopropyl)-2-hydroxy-3-phenylpropanamide (**10**). The mixture of **9** (2.00 g, 6.12 mmol) and NaN₃ (796 mg, 12.24 mmol) in DMSO (20 ml) was stirred at 23° for 4 h. The obtained pale yellow soln. was quenched carefully with H₂O (70 ml, exothermic), allowed to cool, and shaken with Et₂O (50 ml). The aq. layer was extracted with Et₂O (2 × 50 ml), the combined org. soln. washed with H₂O (4 × 15 ml) and sat. aq. NaCl soln. (15 ml), dried (Na₂CO₃), and evaporated, and the residue dried under h.v. (60 min): 1.690 g (95.6%) of pure **10**, White crystalline solid. M.p. 54–64° (dec.). FT-IR (KBr): 3307s, 3240s (NH, OH), 3035m, 2974m, 2926m, 2879m, 2500w, 2100s (N₃), 1961w, 1887w, 1814w, 1643s (C=O), 1633s, 1539s, 1496m, 1455m, 1447m, 1377w, 1353m, 1309s, 1254s, 1194m, 1111s, 1030w, 1003w, 985w, 962w, 948w, 930w, 917w, 841w, 817w, 786w, 732m, 698m, 672w, 651w, 636w, 554m. ¹H-NMR (300 MHz, 20 mg in 0.6 ml of CDCl₃): 7.38 (s, 5 arom. H); 6.41 (br. t, NHCO); 5.08 (d, *J* = 4.0, H–C(3)); 4.46 (br. d, *J* = 4.0, H–C(2)); 3.49 (br. s, OH); 3.30 (m (ddt), *J* = 6.8, 1 H, CH₂NH); 3.09 (q (ddt), 1 H, CH₂NH); 3.05 (dt, ³*J* = 6.7, ²*J* = 12.5, 1 H, CH₂N₃); 2.93 (dt, ³*J* = 6.7, ²*J* = 12.5, 1 H, CH₂N₃); 1.52 (unres. dq_{quint.}, *J* = 0.7, ³*J* = 6.8, CH₂). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.91): 169.64 (CONH); 134.06 (C_{ipso}); 128.85 (C_p); 128.47 (C_m); 128.27 (C_o); 73.96 (C(2)); 67.16 (C(3)); 48.54 (CH₂N₃); 36.24 (CH₂NH); 28.26 (CH₂). CI-MS: 290 (100, [M + H]⁺), 129 (12), 106 (35). Anal. calc. for C₁₂H₁₃N₇O₂ (289.30): H 5.23, C 49.82, N 33.89; found: H 5.53, C 49.75, N 34.12.

(±)-(2R*,3R*)-3-Amino-N-(3-aminopropyl)-2-hydroxy-3-phenylpropanamide (**11**). To a soln. of **10** (1.457 g, 5.042 mmol) in MeOH (50 ml), H₂O (5 ml) and Ph₃P (3.97 g, 15.1 mmol) were added. The obtained suspension was flushed with N₂ and sonicated for 2 h with a gradual temperature increase from 23 to 46°. Evolution of N₂ started at 30° and completely ceased after 60 min to give the expected amount of N₂ (252 ml, 10.8 mmol; no further gas evolution). The clear and colorless soln. was evaporated and the residue partitioned between H₂O (50 ml) and CHCl₃ (50 ml). The aq. layer was extracted with CHCl₃ (5 × 25 ml) and the combined org. soln. extracted with H₂O (25 ml), which was washed with CHCl₃ (5 × 10 ml). The combined org. soln. contained no product. The combined aq. soln. was evaporated at 40° and the residue dried under h.v. at r.t. and then at 60° for 4 h until constant weight: 1.171 g (98%) of pure **11**. Colorless solid. *R*_f (CHCl₃/MeOH/25% aq. NH₃ soln. 10 : 4 : 1, UV and *Fluram* detection) 0.2. M.p. 104–107°. FT-IR (KBr) 3700–2100 (br.), 3348s, 3300s, 3085s, 3063s, 3033s, 2920s, 2869s, 2714s (br.), 2036w, 1950w, 1882w, 1804w, 1647s (C=O), 1583s, 1526s, 1493m, 1455m, 1383w, 1370w, 1350w, 1327m, 1294w, 1270w, 1248w, 1204w, 1185w, 1168w, 1158w, 1120m, 1074m, 1051m, 1013m, 995m, 932m, 913m, 888m, 836w, 816w, 752m, 703s, 619w, 558m. ¹H-NMR (300 MHz, 20 mg in 0.6 ml of (D₆)DMSO, δ(D₂)DMSO = 2.66): 7.66 (t, *J* = 5.7, NHCO); 7.3–7.45 (m, 5 arom. H); 4.23 (d, *J* = 4.6, H–C(2)); 4.16 (d, *J* = 4.6, H–C(3)); 3.21 (sext., *J* = 6.6, 1 H, CH₂NH); 3.11 (sext., *J* = 6.6, 1 H, CH₂NH); 2.48 (unres. td, *J* = 6.6, *J* = 1.2, CH₂NH₂); 1.5–2.5 (very br., OH, NH₂, H₂O); 1.39–1.53 (m, CH₂). ¹³C-NMR (75 MHz, δ((D₆)DMSO) = 39.41): 171.85 (CONH); 142.73 (C_{ipso}); 127.56, 127.15 (C_m, C_o); 126.17 (C_p); 75.81 (C(2)); 57.66 (C(3)); 38.72 (CH₂NH); 35.56 (CH₂NH₂); 32.55 (CH₂). CI-MS: 238 ([M + H]⁺), ESI-MS: 238 (100, [M + H]⁺), 260 (65, [M + Na]⁺). Anal. calc. for C₁₂H₁₉N₃O₂ (237.30): H 8.07, C 60.74, N 17.71; found: H 8.36, C 60.75, N 17.79.

(±)-(2R*,3R*)-2-Hydroxy-3-[[4-(4-methylphenyl)sulfonyl]amino]-N-{3-[[4-(4-methylphenyl)sulfonyl]amino]propyl}-3-phenylpropanamide (**12**). A soln. of **11** (1.146 g, 4.835 mmol) in H₂O (15 ml) was diluted with dioxane (30 ml), Na₂CO₃ (1.025 g, 9.67 mmol) added, and the mixture allowed to cool in an ice/water bath. TsCl (1.90 g, 9.92 mmol) was added by portions and the mixture stirred at 0°, then slowly warming to 20° until no primary amine could be detected by TLC (SiO₂, CHCl₃/MeOH/25% aq. NH₃ soln. 10:4:1; *Fluram*). After 7 h, the mixture was evaporated to a small volume (23°/25 mbar) and quenched with H₂O (70 ml). The crude product was separated by filtration (1.97 g) and crystallized from MeOH/H₂O (75 and 100 ml, resp.): 1.657 g (62.9%) of **12** as white crystalline solid, after washing with H₂O and drying *in vacuo* for 9 h. The product was hardly soluble in all org. solvents, except DMSO and DMF. The combined supernatants were extracted with CHCl₃ to give 560 mg of a residue, which contained no **12** according to TLC and NMR. **12**: *R*_f (5% MeOH/CHCl₃) 0.1. M.p. 150° (dec.) FT-IR (KBr): 3432s, 3331s, 3303s, 3256m, 3065w, 3038w, 2923m, 2872w, 1910w, 1807w, 1756w, 1658s (C=O), 1599w, 1533m, 1495w, 1457m, 1419m, 1322s, 1251w, 1210w, 1184w, 1158s, 1091s, 1065w, 1034w, 1020w, 937w, 809m, 776w, 750w, 703s, 661s, 565s, 550s. ¹H-NMR (300 MHz, 10 mg in 0.6 ml of (D₆)DMSO, δ((D₆)DMSO) = 2.69): 8.27 (*d*, *J* = 9.2, TsNHCH); 7.80 (*d*, *J* = 8.2, 2 H of Ts); 7.67 (*d*, *J* = 8.3, 2 H of Ts); 7.58 (overlapping *d*, *J* = 7.9, 2 H of Ts); 7.55 (overlapping *t*, *J* = 6.6, NHCO); 7.50 (*t*, *J* = 6.1, TsNHCH₂); 7.34 (*d*, *J* = 8.0, 2 H of Ts); 7.15–7.25 (unres. *m*, Ph); 6.11 (*d*, *J* = 5.8, OH); 4.79 (*dd*, *J*(CH,CH) = 3.5, *J*(CH,NH) = 9.2, H–C(3)); 4.22 (*dd*, *J*(CH,CH) = 3.6, *J*(CH,OH) = 5.8, H–C(2)); 3.08, 2.92 (2 *sext.* (*ddt*), *J* = 6.5, CH₂NHCO); 2.58, 2.47 (2 overlapping *s*, 2 Me); 2.58, 2.47 (2 overlapping *m*, OH₂NHTs); 1.38 (*oct.*, *J* = 6.8, CH₂CH₂). ¹³C-NMR (75 MHz, (D₆)DMSO, δ((D₆)DMSO) = 39.41): 170.57 (CONH); 142.36, 141.90, 138.32, 137.57, 136.60 (5 C); 129.44, 128.92, 128.02, 126.92 (4 arom. CH); 126.57 (of Ph); 126.28 (2 overlapping arom. CH); 74.56 (C(2)); 59.14 (C(3)); 39.56 (CH₂NHCO); 35.12 (CH₂NHTs); 28.70 (CH₂); 20.72 (2 Me). ESI-MS (MeCN/CH₂Cl₂): 584 (10, [M + K]⁺), 568 (50, [M + Na]⁺), 546 (100, [M + H]⁺), 375 (53, [M – TsNH]⁺), 255 (24, [OCNH(CH₂)₃NHTs]⁺), 229 (15, [H₃N(CH₂)₃NHTs]⁺). Anal. calc. for C₂₆H₃₁N₃O₆S₂ (545.67): H 5.73, C 57.23, N 7.70, S 11.75; found: H 5.56, C 57.01, N 7.59, S 11.59.

Reaction of 12 with an Excess of 1,4-Dibromobutane (Table 1, Entry 2). A suspension of Cs₂CO₃ (144 mg, 0.44 mmol, 1.2 equiv.) and **12** (200 mg, 0.367 mmol) in anh. DMF (1 ml) was stirred under N₂ for 20 min. Then 1,4-dibromobutane (313 μl, 570 mg, 2.64 mmol, 6 equiv.) was added. After 22 h at 23°, the mixture was partitioned between H₂O (10 ml) and CH₂Cl₂ (3 × 10 ml), dried (Na₂SO₄), and evaporated under h.v. FC (silica gel (25 ml), gradient CHCl₃ → 4% MeOH/CHCl₃) gave disubstituted **18** (39 mg, 12.3%; with 1% MeOH), monosubstituted **16** (81 mg; with 1–2% MeOH), **16/17** 3:1 (63 mg; with 2% MeOH), and unreacted **12** (62 mg, 31%; with 4% MeOH). Yields of **16**, 128 mg (51.3 %) and of **17**, 15.7 mg (6.3%).

Data of (±)-(2R,3R*)-N-{3-[(4-Bromobutyl)](4-methylphenyl)sulfonyl}amino]propyl]-2-hydroxy-3-[[4-(4-methylphenyl)sulfonyl]amino]-3-phenylpropanamide (16)*: White foam. *R*_f (5% MeOH/CHCl₃) 0.26. M.p. 57–59°. FT-IR (KBr): 3000–3700 (br.), 3385s, 3299s, 3064w, 3028w, 2927m, 2870w, 1648s (C=O), 1599m, 1540s, 1495m, 1456s, 1437m, 1331s, 1307m, 1255w, 1185w, 1159s, 1091s, 1063w, 1019w, 973w, 935w, 814m, 703s, 655s, 563s, 549s. ¹H-NMR (300 MHz, 81 mg in 1.3 ml of CDCl₃): 7.59 (overlapping *d*, *J* = 8.1, 2 H of Ts); 7.56 (overlapping *d*, *J* = 8.1, 2 H of Ts); 7.29 (*d*, *J* = 8.1, 2 H of Ts); 7.11–7.18 (*m*, 2 H of Ph); 7.00–7.11 (*m*, 2 H of Ts, 3 H of Ph, NHCO); 6.32 (br. *d*, *J* = 8.1, TsNHCH); 4.82 (unres. *dd*, H–C(3)); 4.55 (*d*, *J*(CH,CH) = 3.6, H–C(2)); 4.4 (very br. *s*, OH); 3.34 (*t*, *J* = 6.5, CH₂Br); 3.24 (*quint.*, 1 H, CH₂NHCO); 2.9–3.1 (*m*, 3 H, CH₂NHCO, CH₂NHTs); 2.71 (br. *t*, *J* = 6.5, Br(CH₂)₂CH₂NHTs); 2.41, 2.29 (2 *s*, 2 Me); 1.76 (*quint.*, *J* = 6.7, CH₂); 1.5–1.6 (overlapping *m*, CH₂); 1.42–1.55, 1.3–1.42 (2 *m*, CH₂). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.92): 170.79 (CONH); 143.37, 143.00 (2 C_p of 2 Ts); 136.98 (C_{ipso} of Ph); 136.06, 135.77 (2 C_{ipso} of 2 Ts); 129.66, 129.23, 128.17, 127.76 (C_o and C_m of Ph, C_m of 2 Ts); 127.45 (C_p of Ph); 127.0 (overlapping, C_o of 2 Ts); 74.51 (C(2)); 59.67 (C(3)), 47.92, 45.53, 35.31, 32.85, 29.51, 28.01, 27.06 (7 CH₂), 21.36, 21.27 (2 Me). ESI-MS: 702, 704 (98, 100, [M + Na]⁺), 680, 682 (17, 17, [M + H]⁺).

Data of (±)-(2R,3R*)-3-[(4-bromobutyl)](4-methylphenyl)sulfonyl]amino]-2-hydroxy-N-{3-[[4-(4-methylphenyl)sulfonyl]amino]propyl}-3-phenylpropanamide (17)*: *R*_f (5% MeOH/CHCl₃) 0.23. ¹H-NMR (300 MHz, CDCl₃; data from **16/17**, selected resonances): 7.69 (*d*, *J* = 8.3, 2 H of Ts); 7.64 (*d*, *J* = 8.4, 2 H of Ts); 5.55 (br. *t*, TsNH); 5.16 (*d*, *J*(CH,CH) = 3.9, H–C(3)); 4.93 (*d*, *J*(CH,CH) = 3.8, H–C(2)). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.90; selected resonances): 172.22 (CONH); 75.13 (C(2)); 63.68 (C(3)).

Data of (±)-(2R,3R*)-3-[(4-bromobutyl)](4-methylphenyl)sulfonyl]amino]-N-{3-[(4-bromobutyl)](4-methylphenyl)sulfonyl]amino]propyl]-2-hydroxy-3-phenylpropanamide (18)*: *R*_f (5% MeOH/CHCl₃) 0.49. ¹H-NMR (300 MHz, 37 mg in 1.0 ml of CDCl₃): 7.66 (overlapping *d*, *J* = 8.3, 2 H of Ts); 7.64 (overlapping *d*, *J* = 8.4, 2 H of Ts); 7.31 (*d*, *J* = 8.0, 2 H of Ts); 7.15–7.29 (*m*, Ph, NHCO, 2 H of Ts); 5.14 (*d*, *J*(CH,CH) = 3.8, CHNTs); 4.84 (*d*, *J*(CH,CH) = 3.8, H–C(2)); 3.2–3.4 (overlapping *m*, 4 CH₂): 3.37 (*t*, *J* = 6.5, CH₂Br); 3.29 (*t*, *J* = 6.5, CH₂); 3.22 (*t*, *J* = 6.5, CH₂); 3.07 (*t*, *J* = 7.3, CH₂); 2.96 (*t*, *J* = 6.6, CH₂); 2.43, 2.40 (2 *s*, 2 Me); 1.76–1.86

(*m*, 2 H); 1.4–1.75 (*m*, 8 H). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.90): 171.04 (CONH); 143.37 (2 C_p of 2 Ts); 137.33 (C_{ipso} of Ph); 136.18, 135.08 (2 C_{ipso} of 2 Ts); 129.69, 129.60, 129.41, 128.16 (C_o and C_m of Ph, C_m of 2 Ts); 128.06 (C_p of Ph); 127.15, 127.00 (C_o of 2 Ts); 74.58 (C(2)); 64.42 (C(3)); 48.06, 47.37, 45.78, 35.78 (4 CH₂); 32.81 (br. *s*, 2 overlapping CH₂); 29.85, 29.52 (2 CH₂); 28.24 (br. *s*, 2 overlapping CH₂); 27.13 (CH₂); 21.37 (br. *s*, 2 overlapping Me). ESI-MS: 836, 838, 840 (50, 100, 50, [M + Na]⁺), 814, 816, 818 (5, 10, 5, [M + H]⁺).

Macrocyclization of 12 with 1,4-Dibromobutane under High Dilution (Table 1, Entry 4). A soln. of **12** (801 mg, 1.470 mmol) and 1,4-dibromobutane (192 μl, 1.635 mmol) in DMF (250 ml) was added dropwise to a stirred suspension of Cs₂CO₃ (1.006 g, 3.087 mmol) in DMF (500 ml) at 60° under N₂ within 8 h. After 24 h at 60°, additional 1,4-dibromobutane (192 μl, 1.635 mmol) and Cs₂CO₃ (513 mg) were added, and stirring was continued for another 24 h. The mixture was quenched with H₂O (24 ml) and AcOH (0.5 ml, pH 7) and evaporated at 45°/12 mbar. The crude brown oil was partitioned between CH₂Cl₂ (4 × 50 ml) and H₂O (100 ml), and the combined org. soln. dried (Na₂SO₄) and evaporated to give 1.182 g of brown oil. FC (silica gel (100 ml), CHCl₃ → 5% MeOH/CHCl₃) gave dicarbamate **23** (83 mg; with CHCl₃), formimidamide **21** (46 mg, 13.8%; with CHCl₃ and 1% MeOH), macrocyclic **13** (167 mg, 19%; with 1% MeOH), carbamate **22** (132 mg; with 2% MeOH); carbamate **20** (73 mg, 7.2%; with 2% MeOH), starting **12** (247 mg, 30%; with 3% MeOH), and finally hydroxy compound **19** (45 mg, 5%; with 5% MeOH).

Data of (±)-(2R*,3R*)-3-Hydroxy-1,9-bis[(4-methylphenyl)sulfonyl]-2-phenyl-1,5,9-triazacyclotridecan-4-one (13): White solid. M.p. 176–187° (dec.). R_f (5% MeOH/CHCl₃) 0.32. Complete ¹H- and ¹³C-NMR assignments from 2D NMR at 60° (at r.t. many ¹H- and ¹³C-NMR signals were broad), see Table 3. ¹H-NMR (300 MHz, 60 mg in 0.7 ml of CDCl₃, 23°): 7.69 (*d*, *J* = 8.0, 2 H of Ts–N(9)); 7.40 (br. *d*, *J* = 6.4, 2 H of Ts–N(1)); 7.32 (*d*, *J* = 8.0, 2 H of Ts–N(9)); 7.10–7.20 (*m*, Ph, NHCO); 6.96 (br. *d*, *J* = 6.7, 2 H of Ts–N(1)); 4.94 (br. *d*, H–C(2)); 4.73 (br. *s*, H–C(3)); 3.76 (*m*, 1 H, CH₂N); 2.95–3.50 (overlapping br. *m*, 7 H, CH₂N); 2.43, 2.36 (2 *s*, 2 Me); 1.5–1.69, 1.69–1.87, 1.87–2.02, 2.02–2.2 (overlapping br. *m*, 3 CH₂). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.97, 23°): 171.68 (br. *s*, CONH); 143.54, 143.35 (2 C_p of 2 Ts); 136.74 (br. *s*, C_{ipso} of Ph); 134.91 (br. *s*, C_{ipso} of Ts–N(1)); 134.61 (C_{ipso} of Ts–N(9)); 129.70, 129.29, 128.89, 128.17 (C_o and C_m of Ph, C_m of 2 Ts); 127.89 (C_p of Ph); 127.35 (overlapping C_o of 2 Ts); 71.16 (br. *s*, C(3)); 65.11 (br. *s*, C(2)); 50.66, 46.56, 39.06, 28.13, 27.2, 25.80 (6 br. *s*, 7 CH₂); 21.39 (br. *s*, 2 Me). ESI-MS: 622 (100, [M + Na]⁺).

Data of (±)-(2R*,3R*)-2-Hydroxy-N-[3-[(4-hydroxybutyl)[(4-methylphenyl)sulfonyl]amino]propyl]-3-[[4-methylphenyl)sulfonyl]amino]-3-phenylpropanamide (19): R_f (5% MeOH/CHCl₃) 0.07. ¹H-NMR (300 MHz, 6 mg in 0.5 ml of CDCl₃): 7.61 (*d*, *J* = 8.3, 2 H of Ts); 7.55 (*d*, *J* = 8.3, 2 H of Ts); 7.30 (*d*, *J* = 8.0, 2 H of Ts); 7.05–7.15 (*m*, 7 arom. H); 7.00 (br. *t*, *J* = 6.3, NHCO); 6.05 (br. *s*, NHTs); 4.72 (unres. *dd*, H–C(3)); 4.47 (*d*, J(CH,CH) = 4.32, H–C(2)); 3.62 (*t*, *J* = 5.7, CH₂OH); 3.10–3.22 (*m*, CH₂NHCO); 2.92–3.12 (*m*, CH₂NTs); 2.76–2.84 (unres. *m*, CH₂NTs); 2.42, 2.32 (2 *s*, 2 Me); 1.35–1.65 (*m*, 3 CH₂). ¹³C-NMR (75 MHz, 45 mg in 0.6 ml CDCl₃, δ(CDCl₃) = 76.93): 171.15 (CONH); 143.26, 142.90 (2 C_p of 2 Ts); 137.06, 135.93, 135.70 (C_{ipso} of Ph, C_{ipso} of 2 Ts); 129.62, 129.17, 128.17, 127.67 (4 arom. CH); 127.35 (C_p of Ph); 127.01 (2 overlapping arom. CH); 74.60 (C(2)); 61.88 (CH₂OH); 59.52 (C(3)); 49.01 (CH₂NTs); 45.79 (CH₂NTs); 35.57 (CH₂NHCO); 29.39 (CH₂); 28.23 (CH₂); 25.28 (CH₂); 21.36, 21.26 (2 Me). ESI-MS: 618 (7, [M + H]⁺), 640 (100, [M + Na]⁺).

Data of (±)-4-[[3-[(2R*,3R*)-2-Hydroxy-3-[[4-methylphenyl)sulfonyl]amino]-1-oxo-3-phenylpropyl]amino]propyl]-(4-methylphenyl)sulfonyl]amino]butyl Dimethylcarbamate (20): R_f (5% MeOH/CHCl₃) 0.2. ¹H-NMR (300 MHz, 74 mg in 0.8 ml of CDCl₃): 7.56 (*d*, *J* = 7.9, 2 H of Ts); 7.52 (*d*, *J* = 8.0, 2 H of Ts); 7.26 (*d*, *J* = 7.7, 2 H of Ts); 6.95–7.15 (*m*, 7 arom. H, NHCO); 6.38 (br. *d*, NHTs); 4.80 (br. *s* (unres. *dd*) CHNHTs); 4.48 (br. *d*, *J* = 3, CHOH); 3.95–4.01 (unres. *m*, CH₂OCON); 3.15–3.28 (br. *m*, 1 H); 2.9–3.1 (br. *m*, 3 H); 2.85 (*s*, Me₂N); 2.6–2.7 (br. *s* (unres. *m*), CH₂NTs); 2.39, 2.27 (2 *s*, 2 Me); 1.4–1.6 (br. *m*, 5 H, CH₂); 1.25–1.35 (br. *m*, 1 H, CH₂). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.97): 170.80 (CONH); 156.63 (Me₂NC(O)OR); 143.23, 142.83 (2 C_p of 2 Ts); 137.19, 136.14, 135.91 (C_{ipso} of Ph, C_{ipso} of 2 Ts); 129.60, 129.16, 128.22, 127.62 (4 arom. CH); 127.30 (C_p of Ph); 126.97 (2 overlapping arom. CH); 74.53 (CHOH); 64.58 (CH₂OCON); 59.55 (CHNHTs); 48.61 (CH₂); 45.50 (CH₂); 36.26, 35.80 (2 br. *s*, Me₂N); 35.17 (CH₂); 28.13 (CH₂); 26.22 (CH₂); 25.23 (CH₂); 21.36, 21.26 (2 Me of 2 Ts). ESI-MS: 711 (100, [M + Na]⁺), 689 (5, [M + H]⁺).

Data of N-[(Dimethylamino)methylene]-4-methylbenzenesulfonamide (21) [23]: R_f (5% MeOH/CHCl₃) 0.5. ¹H-NMR (300 MHz, CDCl₃): identical with [23]; 8.13 (*s*, NH); 7.77 (*d*, *J* = 8, 2 H of Ts); 7.25 (*d*, *J* = 8, 2 H of Ts); 3.12, 3.01 (2 *s*, Me₂N); 2.40 (*s*, Me of Ts). ¹³C-NMR (75 MHz, δ(CDCl₃) = 76.97): 159.04 (HC = NTs); 142.30, 139.53 (C_p and C_{ipso}, resp., of Ts); 129.19 (2 H_m of Ts); 126.39 (2 H_o of Ts); 41.39, 35.41 (Me₂N); 21.33 (Me of Ts).

Data of 4-Hydroxybutyl Dimethylcarbamate (22): R_f (5% MeOH/CHCl₃) 0.23. ¹H-NMR (300 MHz, CDCl₃): 4.10 (*t*, *J* = 6.3, CH₂OCON); 3.67 (*t*, *J* = 6.2, CH₂OH); 2.90 (*s*, Me₂N); 1.6–1.8 (*m*, CH₂CH₂). ¹³C-NMR

(75 MHz, $\delta(\text{CDCl}_3) = 76.92$): 156.70 ($\text{Me}_2\text{NC}(\text{O})\text{OR}$); 65.02 (CH_2OCON); 62.25 (CH_2OH); 36.17 (br. s, Me_2N); 28.99, 25.48 (CH_2CH_2).

Data of 1,4-Butanediyl Dimethylcarbamate (23): R_f (5% MeOH/ CHCl_3) 0.6. $^1\text{H-NMR}$ (300 MHz, CDCl_3): 4.07–4.13 (unres. m, 2 CH_2O); 2.90 (s, 2 Me_2N); 1.70–1.88 (unres. m, 4 H). $^{13}\text{C-NMR}$ (75 MHz, $\delta(\text{CDCl}_3) = 76.94$): 156.54 ($\text{Me}_2\text{NC}(\text{O})\text{OR}$); 64.82 (CH_2O); 35.73, 35.36 (2 br. s, Me_2N); 25.70 (CH_2).

Reaction of 12 with 1,4-Dibromobutane under Moderate Dilution (Table 1, Entry 1): A mixture of **12** (30.2 mg, 0.0554 mmol), 1,4-dibromobutane (14.4 mg, 0.0665 mmol), and Cs_2CO_3 (45.1 mg, 0.139 mmol) in anh. DMF (5 ml) was stirred at 23° under N_2 for 44 h. The resulting mixture was partitioned between CHCl_3 (30 ml) and 5% aq. citric acid (2 ml) and the org. phase washed with H_2O (5×5 ml), dried (Na_2SO_4), and evaporated under h.v.: 41 mg of white solid. FC/silica gel (10 ml, $\text{CHCl}_3 \rightarrow$ 5% MeOH/ CHCl_3) gave **13** (3 mg, 9%), **16** (21 mg, 56%), and **24** (ca. 1:1 mixture of 2 diastereoisomers; 8 mg, 25.2%, with 3% MeOH).

Data of (\pm)-(2R,3R*)-N,N'-[1,4-Butanediylbis[[(4-methylphenyl)sulfonyl]imino]-3,1-propanediyl]-bis[2-hydroxy-3-[(4-methylphenyl)sulfonyl]amino]-3-phenylpropanamide] (24):* $^1\text{H-NMR}$ (300 MHz, 8 mg in 0.6 ml of CDCl_3): 7.60 (d, $J = 8.3$, 4 H of Ts); 7.53 (d, $J = 7.85$, 4 H of Ts); 7.29 (d, $J = 7.7$, 4 H of Ts); 6.95–7.12 (m, 12 H, Ph, Ts, 2 NHCO); 6.25–6.42 (2 very br. s, 2 H, disappeared after D_2O exchange, 2 TsNHCH); 4.80 (br. s, 2 H, became 2 d at 4.81 ($J(\text{CH},\text{CH}) = 4.2$) and 4.78 ($J(\text{CH},\text{CH}) = 4.3$) after D_2O exchange, 2 PhCHNHTs); 4.56 (d, $J = 4.15$, 1 H, 2 CHOH of one diastereoisomer); 4.51 (d, $J = 4.25$, 1 H, 2 CHOH of the other diastereoisomer); 2.85–3.15 (m, 12 H); 2.78 (m, 4 H); 2.42 (s, 6 H, Me); 2.30, 2.29 (2 s, 6 H, 2 Me); 1.34–1.65 (m, 12 H). ESI-MS: 595 (100, $[\text{M} + 2\text{Na}]^{2+}$).

Intramolecular Macrocyclization of 16. A soln. of **16** (78 mg, 0.115 mmol) in DMF (30 ml) was stirred in the presence of Cs_2CO_3 (90 mg, 0.276 mmol) under N_2 for 75 h at 40° and then evaporated. The residue was separated by FC as described above to give **13** (20 mg, 29.1%), **21** (2 mg, 7.7%), unreacted **16** (15 mg, 19%), **20** (10 mg, 10.2%), and **21** (6 mg, 8.5%).

Macrocyclization of 12 with 1,4-Dibromobutane at Moderate Dilution (Table 1, Entry 5). A soln. of **12** (42 mg, 0.0771 mmol) and 1,4-dibromobutane (18.3 mg, 0.0848 mmol) in anh. DMF (5 ml) was stirred in the presence of Cs_2CO_3 for 24 h at 50° under N_2 . The mixture was quenched with a small amount of AcOH, evaporated under h.v., and partitioned between CHCl_3 and H_2O . Macrocyclic **13** (10.0 mg, 21.7%) was isolated after FC (silica gel, 1% MeOH/ CHCl_3).

Electrochemical Deprotection of Tosyl Groups [25]: (\pm)-(2R*,3R*)-3-Hydroxy-2-phenyl-1,5,9-triazacyclotridecan-4-one (**14**). Controlled-potential electrolysis was carried out in a cylindrical, three electrode, divided cell with an electronic potentiostat and a stirred mercury pool (area: 44 cm^2) as cathode, graphite rod as anode, and SCE as reference electrode. First, 100 ml of 0.1M (Me_4N)Cl in 94% EtOH (used as catholyte and anolyte) were electrolyzed at -2.25 V under Ar at $+5^\circ$ until the background current remained constant (10 mA). Then **13** (100 mg, 0.167 mmol) in a minimum amount of DMF was added to the cathodic chamber. The electrolysis was carried at -2.25 V until current depletion to the background level was observed in the recorded I/t curve. Approximately 400% of the necessary number of coulombs were used. The cathodic soln. was then evaporated, the solid residue dissolved in H_2O (5 ml), saturated with K_2CO_3 , and extracted with CH_2Cl_2 (5×10 ml). The combined org. phase was dried (Na_2CO_3) and evaporated to yield 75 mg of crude **14** contaminated with traces of electrolyte (Me_4N)Cl, potassium toluenesulfonate, and DMF. The substance was used in the next step without further purification. R_f ($\text{CHCl}_3/\text{MeOH}/25\%$ aq. NH_3 soln. 78:19:3) 0.23. CI-MS: 292 (100, $[\text{M} + \text{H}]^+$). $^1\text{H-NMR}$ (300 MHz, 75 mg in 0.8 ml of CDCl_3 , see also Table 3): 7.13–7.28 (m, Ph, NHCO); 3.98 (d, $J = 9.1$, H–C(3)); 3.56 (d, $J = 9.0$, H–C(2)); 3.37–3.49 (m, 1 H, CH_2NH); 3.22–3.34 (m, 1 H, CH_2NH); 2.68–2.80 (m, 2 H); 2.55–2.60 (m, 2 H); 2.38–2.48 (m, 1 H); 2.21–2.31 (m, 1 H); 1.63–1.73 (m, 2 H); 1.53–1.68 (m, 3 H); 1.30–1.42 (m, 1 H). $^{13}\text{C-NMR}$ (75 MHz, $\delta(\text{CDCl}_3) = 77.05$, see also Table 3): 173.13 (CONH); 141.00 (C_{ipso} of Ph); 128.71, 128.21, 127.71 (C_o , C_m , and C_p of Ph); 73.6 (C(3)); 66.44 (CHNH); 49.50, 48.82, 45.78, 39.60 (4 CH_2N); 27.86, 27.81, 27.52 (3 CH_2).

(\pm)-(2R*,3R*)-3-Hydroxy-9-[2(E)-1-oxo-3-phenylprop-2-enyl]-2-phenyl-1,5,9-triazacyclotridecan-4-one (= 3-Hydroxycelaccinnine; **1**). According to the procedure of Yamamoto and Maruoka described for celaccinnine ($\text{Et}_3\text{N}/\text{DMAP}$, (N,N -dimethylpyridin-4-amine); -78°) [26], **14** was monoacylated. The product was purified by two FC (silica gel, first 2% MeOH/ CHCl_3 (**1** eluted together with DMAP), then 8% MeOH/ CHCl_3): 20 mg (33% from **13**, not optimized) of **1**. R_f (10% MeOH/ CHCl_3) 0.35. White solid. M.p. 146–150° (dec.). FT-IR (KBr): 3296s (br., OH, NH), 3060w, 3026w, 2927s, 2855m, 1758w, 1731w, 1647s (C=O), 1595s, 1542m, 1496m, 1496m, 1452m, 1435s, 1377w, 1359w, 1328m, 1310m, 1245w, 1203m, 1121m, 1084w, 1054w, 985w, 911w, 854w, 762m, 702s. NMR: see Table 2 for complete signal assignments from 2D NMR data at $+70^\circ$ and -40° . $^1\text{H-NMR}$ (300 MHz, 16 mg in 0.6 ml of CDCl_3 , 23°): 8.36–8.48 (br. s, 0.5 H, NHCO); 7.93–8.05 (br. s, 0.5 H, NHCO); 7.70 (d, $J_{\text{trans}} = 15.4$, PhCH=CH); 7.48–7.56 (m, 2 H of PhCH=CH); 7.28–7.43 (m, 8 arom. H); 6.80 (d, $J_{\text{trans}} = 15.4$,

1 H, PhCH=CH); 4.08 (*d*, $J(\text{CH}, \text{CH}) = 9.6$, 1 H, CHOH); 3.90–4.05 (br. *m*, 0.5 H); 3.42–3.75 (overlapping *m*, 4 H); 3.61 (*d*, $J(\text{CH}, \text{CH}) = 9.60$, PhCHNH); 3.22–3.42 (br. *m*, 2 H); 3.05–3.25 (br. *s*, 0.5 H); 2.62–2.72 (*m*, 1 H); 2.42–2.52 (*m*, 1 H); 2.0–2.2 (br. *m*, 1 H); 1.85–2.1 (br. *m*, 2 H); 1.65–1.85 (br. *m*, 2 H); 1.4–1.65 (br. *m*, 3 H). CI-MS: 422 (100, $[M + H]^+$).

(±)-(2R*,3R*)-3-Hydroxy-9-[4-methylphenyl)sulfonyl]-2-phenyl-1,5,9-triazacyclotridecan-4-one (**15**).

The procedure and data will be provided in a separate publication [24]. For the NMR data, see Table 3.

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